

■ *Heterobostrachus aequalis*

Waterhouse

BIOLOGY

This is a polyphagous species and a common pest of packing cases, plywood chests and other wood-based industrial products. It is a minor pest of dry bamboos. The adult beetle is black, 6-15 mm long, cylindrical and with a rough hooded prothorax, usually with curved hooks at the distal ends of elytra. Female adult lays eggs singly on rough surfaces of dry bamboos. The larvae, after hatching, bore into the soft portion and gradually make a wide tunnel. The tunnels are tightly packed with fine wood dust and the host bamboo is soon reduced to powder. The life cycle of the beetle is annual.

Other bostrychid beetles commonly seen are *Dinoderus pilifrons* Lesne, *Cryphalus satonis* Mat. and *Heterobostrychus hamatipennis* Lesne.

Powder-post Beetles (Coleoptera: Lyctidae)

Several lyctid beetles primarily attack dried bamboo timber and bamboo products. Although these species are quiet common, they cause much less damage to bamboos than bostrychid beetles.

■ *Lyctus africanus* Lesne

DISTRIBUTION

India.

HOST

Bambusa bambos and *Dendrocalamus strictus*.



BIOLOGY

The beetle is flat, light brown to almost black in colour, and about 2-7 mm long. After hatching, larvae start feeding on wood tissues converting it to a very fine

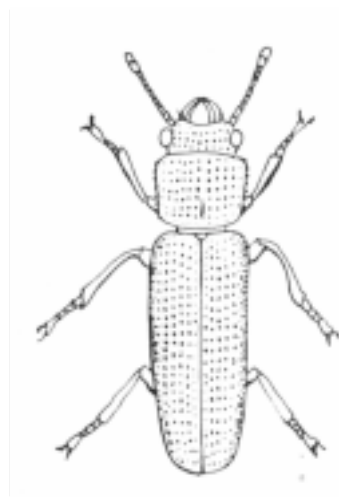


Fig. 58: Adult of *Lyctus africanus*

powder, which remains closely packed in the tunnel. Moisture content of less than 10% is unsuitable for development of the larvae. After about one month of pupal period, beetles emerge by cutting approximately circular holes. The life-cycle varies from one to three years, depending on the availability of food and moisture within the wood.

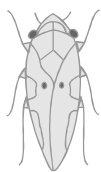
CONTROL

Spraying with lindane 0.1% in kerosene is recommended (Sandhu 1974).

■ *Lyctus brunneus* Stephens

DISTRIBUTION

Tropical and subtropical regions, particularly in Southeast Asian countries.



BIOLOGY

This is a major boring pest on timber of various broadleaf tree species and woody products, but cause only minor damage to bamboo. The adult beetle is 2.2-7 mm long and varies from light brown to reddish-brown in colour. The protergum of the adult is rectangular, with a clearly longitudinal, Y-shaped suture on it. Both adults and larvae bore coaxially to the culm fibre. There is one generation per year. Adults emerge throughout the spring and summer months. Larvae occur from May to the following March and pupate in tunnels when fully grown (Chen Liang 1983).

Other Powder-post Beetles

The Chinese powder-post beetle, *Lyctus sinensis* Lesne, is widespread in tropical and subtropical areas, especially in Southeast Asian countries. It attacks timber of many trees, including bamboos. The adult beetle is 3-6 mm long, 0.8-1.2 mm wide and dark brown in colour. The protergum of the adult is oblong and the longitudinal suture on it is not prominent. This beetle has an annual life cycle and overwinters as larvae in tunnels. Both adults and larvae bore in dried culms and finished products, and the damaging process can last for over eight years. *Tillus notatus* Klug, *Tarsostenus univittatus* Rossi and *Chelifer* sp. prey on the larvae and pupae of the borer (Zhan Zhongcai 1983).

Minthea rugicollis Walker is widespread in tropical and subtropical regions. The adult is 2.3-2.8 mm long and covered with greyish-white hair. The protergum is concave in the centre. There are three generations per year. Both adults and larvae feed on dried wood materials throughout the year, but are relatively less active in the winter season.



Most of the measures for controlling bostrychid beetles are also effective for these lyctid borers.

Long-horned Beetles (Coleoptera: Cerambycidae)

■ Grey long-horned beetle - *Niphona furcata* Bates

DISTRIBUTION

China and Japan.

HOSTS

Newly felled culms, which are less than 2 cm in diameter, of *Phyllostachys* and *Pleioblastus* species.

BIOLOGY

Adult beetles are 13-18 mm long, 4-4.5 mm wide and varies from grey to dark brown in colour. There is one generation per year, overwintering as adults in the tunnels bored. Adults come out in April and feed on young twigs, newly-sprouted leaves and culm tissue. Female adults make splits on culms selected for egg-laying and deposit eggs into culm cavities. Eggs are laid individually, with only one egg per culm in most cases. Larvae, occurring from May to October, feed on the inner surface of the culm, making shallow tunnels, and repeated feeding causes the damaged culm to thin down and become susceptible to collapse in wind (Zhang Xiankai and Zuo YX 1988).

■ Tiger longicorn - *Chlorophorus annularis* Fabricius

DISTRIBUTION

China, India, Indonesia, Japan, Malaysia, Myanmar, Thailand and Vietnam.



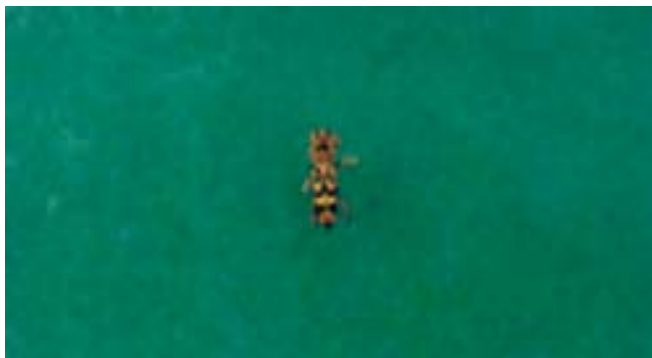


Fig. 59: Adult of *Chlorophorus annularis*

HOSTS

Primarily on felled *Phyllostachys* spp., *Bambusa* spp. and *Dendrocalamus* spp. culms, but also on apple, Chinese sweet gum, teak and cotton tree trunks.

BIOLOGY

The adult beetle is 9.5-17 mm long, 2.4-2.5 mm wide and ochre yellow in colour, with dark brown or black curved markings on the elytra and pronotum. Normally, there is one generation per year. However, larval development rate is affected by the dryness of the culm, and may continue even after the culm has been made into products. Adults occur from April to September and eggs are laid on cut ends or in cracks. Larvae overwinter in tunnels in the culm.

■ *Ceresium sinicum* White

DISTRIBUTION

China, Japan and Thailand.

BIOLOGY

This is an important wood borer, the larvae of which attack felled logs of various trees, including bamboos. The adult beetle is 10-15 mm long, 2-3.5 mm wide



and brown or light brown in colour. The protergum is irregularly marked, but has a smooth, longitudinal ridge (carina) at its centre. There is one generation per year. Adults emerge in May, and eggs are deposited onto culm tissue through wounds and cracks. Larvae bore in the culm wall, forming vertical tunnels, and overwinter in the tunnel when fully grown. *Paracerchysius ceresii*, *Zombrus bicolor* and *Scleroderma* sp. parasitize larvae and pupae of the beetle (Zhan Zhongcai 1986).

■ Other Long-horned Beetles

The teak trunk borer, *Stromatium barbatum* Fab., is primarily a pest of packing cases, seasoned timbers, furniture, plywood, and wood work in building. It also attacks bamboos. The beetle deposits eggs in small holes on rough surfaces of the host wood. The larval tunnels are cylindrical and tightly packed with fine wood dust, which is normally ejected out at intervals.



Fig. 60: Adult of *Stromatium barbatum*



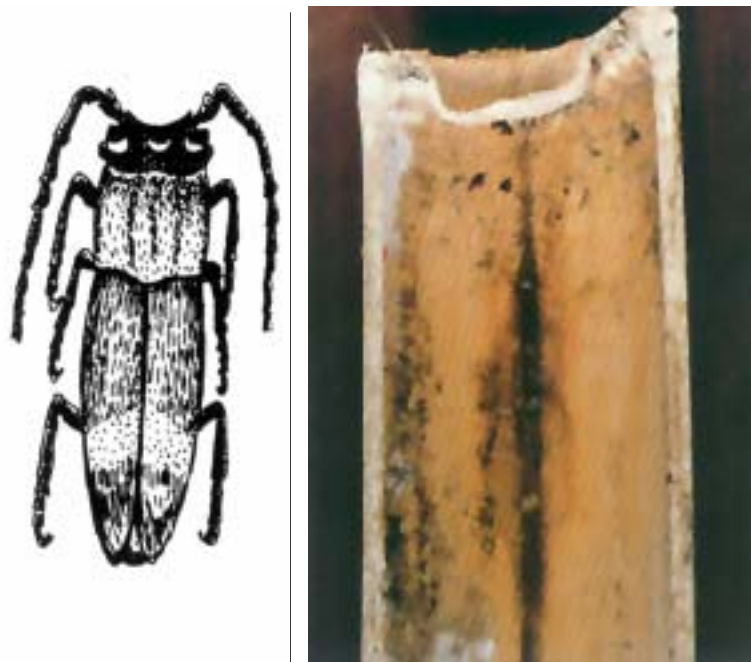


Fig. 61: Adult of *Diboma posticata* and the damage caused

The larvae produce distinctly audible sound of scraping of wood with mandibles.

Other common long-horned beetles attacking post-harvest bamboo include *Chlorophorus figuratus* Scop, *Glenea notata* Gahan, *Niphona hookeri* Gahan, *Diboma posticata* Gahan, *Merionoeda nigriceps* White, *Purpuricenus sanguinolentus* Oliver and *Ropica signata* Pic.

Termites

The American powder-post termite, *Cryptotermes dudleyi* Banks (Isoptera: Kalotermitidae), is an introduced species found in Bangladesh and India (restricted to coastal regions of Orissa, West Bengal and Daman). The attack is usually serious, and the



infested material gets completely riddled with excavations, to be ruined within a short period. All types of bamboo constructions, including split bamboos used as rafters or matted walls in houses, are seriously attacked. The attack by this species resembles a typical bostrychid attack with small holes appearing on the outer portion (Chaudhry 1955; Sen-Sarma and Mathur 1957; Sen-Sarma et al. 1975; Thakur 1988a).

Odontotermes feae Wasm. (Isoptera: Macrotermitidae) is an important wood-destroying termite species attacking wood and bamboo structures, bamboo poles and matted or split bamboos in Bangladesh, India and Myanmar. This species builds narrow, covered earthen runways or broad sheaths of mud plaster on the surface of felled or cut bamboos on the ground (Beeson 1941; Bhasin et al. 1958).

Coptotermes heimi Wasm. (Isoptera: Rhinotermitidae) is a highly polyphagous species with as many as 35 species of host plants, in addition to bamboos. It is found associated with bamboo in India and Pakistan. The pest eats away the inner soft portion along the fibre, leaving the outer surface intact. In seriously infested bamboos, the inner portion looks like a honeycomb.

Microcerotermes fletcheri Holmg. (Isoptera: Rhinotermitidae) is found occasionally in dead bamboo clumps, making numerous honeycomb cavities (Beeson 1941; Roonwal 1970).

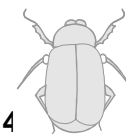


9. INTEGRATED INSECT PEST MANAGEMENT

Over 800 insect species have been reported as bamboo pests in the Asian region. The damage they cause and the threat they pose to bamboo-based industries have been identified in some countries as more large-scale, intensively managed plantations are being established. Various control measures against outbreaks of insect pests have been developed, but most of these involve the application of chemical insecticides. Besides polluting the environment, the use of large amounts of chemical insecticides also removes the natural enemies of pests, thereby weakening natural control systems and shortening the interval of outbreaks. Therefore, the long-term plan and related strategies that address insect pest problems on bamboo should be based on integrated management measures. Integrated pest management (IPM) is the comprehensive approach that considers all available measures as a whole to solve pest problems. Although an IPM program for bamboo is not readily available yet, investigations are being undertaken and the approaches will include silvicultural, biological, direct and chemical controls.

Silvicultural Control

Silvicultural control, the first step of IPM, combines all possible cultural measures to ensure that the plants are exposed to the least possible pest-stress. When managed properly, silvicultural control can be an effective way to minimize insect damage and hence, is considered primary to IPM.



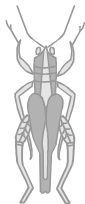
Silvicultural control of pest insects in bamboo stands is achieved mainly through proper tending, regulating culm density and age structure, and regenerating the stands. These measures aim to kill pest insects directly, create unfavourable environment conditions for the pests and improve the insect-resistant ability of the plants.

Soil-turning, an important measure of bamboo cultivation, can significantly improve nutrient and water supply conditions at the site and enhance the vigour of bamboo plants. If undertaken in winter months, it is effective in suppressing the population of those species already living, overwintering or diapausing in the soil, either by directly killing or by exposing them to unfavourable weather conditions and natural enemies.

Direct Control

This approach to pest control targets the reduction of pest population through mechanical or physical means. Aiming to prevent numerical build-up of insects or reduce the damage caused by them, direct control measures in bamboo stands often employ removing, trapping and setting physical barriers to pests. These measures are designed according to common or specific behaviour of target insects and can be very effective though labour-intensive. Direct control approach needs a detailed understanding of the biology and life history of the target insect pests.

Adults of a number of insect pest species attacking bamboos, especially those in the lepidopteran species, are active at nights, displaying strong phototaxis. Light-trapping at nights during adult stage can significantly reduce larval density. Mass-trapping in this manner has proven practical and successful in



suppressing the important bamboo leaf feeders in China. Devices required for this are simple, and a choice of types is available to suit the insect species involved, size of stands and user preferences. Black fluorescent light lamp is the one employed most commonly. The lamp can be set at an open place nearby or in the stand. It is kept about 1-1.5 m above the ground, and a container with an aqueous insecticide solution is placed below the lamp, or an electric fence is set beside or around the lamp to kill the attracted insects. Best results are obtained in dark, hot and windless nights.

Some insects that feed on bamboo are attracted to a particular food or odour. Such behavioural responses, referred to as chemotaxis, can also be used in controlling the pests. Toxic baits or traps can be very effective in suppressing the population of such insects. Both adults and nymphs of bamboo locusts, for example, prefer salty food and uric odour, and mass trapping can be achieved using pots containing a mixed solution of bitter and insecticides. Similarly, large number of adults of all bamboo shoot fly species can be captured and killed by traps baited with sweet and sour materials, or using fresh bamboo shoot pieces treated with insecticides. In case of bamboo leaf-rollers, adult moths need to feed on flower juice and show a strong preference to oak and chestnut trees. As a long-term measure, some of these trees can be planted to serve as traps, and satisfactory control can be achieved by spraying the trees with insecticides when the moths gather on them in swarms to feed.

Setting physical barriers to deter insects from reaching the point of attack is commonly used to control forest insect pests. It is particularly effective against insect species that crawl to reach host plants. As far as

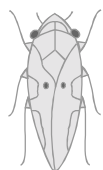


bamboo is concerned, the most successful use of this method is in the control of the stink bug, *Hippotiscus dorsalis*, which feeds on culms and branches. The bug has an annual life cycle and overwinters as nymphs on the ground. The wingless nymphs move onto culms by crawling in the spring season. Setting sticky or adhesive barriers around the basal part of culms in early spring can prevent the overwintering nymphs from reaching the culm. The bugs accumulated on culms under the barrier can be killed using methods such as spraying contact insecticides.

Removal of insects from their habitat and host plants is the simplest way among direct control measures. It is very effective in some cases to alleviate pest pressure on bamboo plants and practical in small-scale stands or isolated plantings. The damage caused by bamboo shoot weevils can be significantly reduced by removing adult weevils from bamboo shoots by hand-picking before any egg-laying occurs, because adult weevils of these species will drop to the ground when disturbed. Eggs of tussock moth are mainly deposited in groups at the basal part of culms, and can easily be removed or destroyed to significantly reduce larvae population. Some other insects, which feed in groups at the basal portion of culms or on leaves of lower branches, can also be physically removed.

Biological Control

Almost all insect pests are attacked by a number of other living organisms, which are their natural enemies. The use of such organisms to suppress population density of insect pests and the damage they cause is referred to as biological control, and has become an important IPM component. The natural enemies of insect pests are often grouped as parasitoids, predators and pathogens. Parasitoids are insect species that



parasitize other insects. Majority of them and the most important species are hymenopteran wasps and dipterian flies. They feed as larvae on host insects during egg, larva or pupa stages, and cause the death of the hosts when they complete their own development of larval stage. Predatory natural enemies of insect pests include a variety of mammals, birds, spiders and other insects. Various species of bacteria, viruses and fungi are pathogenic to insect pests. The use of insect pathogens and their products has become a major tool for biological control.

In natural bamboo stands, the bamboo eco-system remains mostly intact as biodiversity and natural enemies of pests play important roles in regulating insect pest populations. However, when bamboos are cultivated intensively, such as in large-scale pure plantations, biodiversity hardly exists. Besides, complete weeding and use of large amounts of chemical insecticides destroy the habitat and food resources of natural enemies and the natural control reduces sharply. Inundative release of natural enemies to control insect pests on bamboos has been practised successfully in some countries, but this method does not offer long-term control. Conservation of natural enemies is an important strategy of biological control aiming to sustain environmental conditions favourable to natural enemies. Measures such as retaining the surrounding vegetation and reducing the use of chemical insecticides form parts of such a strategy.

Chemical Control

Chemical control involves mainly the use of insecticides to kill insect pests. Judiciously used, chemical control is highly effective, easy to use and



low in cost. It is an important approach and the most widely used in insect pest management. A number of insecticides are available, varying in structure, formulation and action. The choice of insecticide is made according to the insect and host species involved, available equipment, and application techniques.

Insecticides can be applied to standing bamboo plants by dusting, spraying, injecting and smoking. Dusting with insecticide is an economical way for controlling bamboo leaf feeders. Mist-spraying — emulsifiable concentrates or wettable powders diluted in water and applied using mist blowers — is the most widely used technique and is effective against almost all leaf feeders and some sap suckers on bamboos. Smoking is recommended for controlling bamboo leaf feeders and some sap suckers in stands which are high in density or located in mountainous areas. It is important that smoking be done in windless weather, usually in the early morning or later afternoon. Systemic insecticides can be absorbed and transmitted by plant tissues and hence, has an important place in pest management in forests. The application of systemics by injecting into the culm cavity or applying to the root system has been particularly recommended and, because it is relatively safe to natural enemies and environments, it is being widely used in China for controlling insect pests — including leaf feeders, sap suckers, shoot borers and gall makers — on monopodial bamboos with large-sized culms.

It is well known that many chemical insecticides are broad-spectrum and hence, highly toxic to mammals and natural enemies. Repeated use of large amounts of chemical insecticides also induces resistance in insects and results in environmental hazards. Chemical control is to be considered only when other management



measures are not capable of preventing pest outbreaks. Selection of the correct insecticide, application method and timing play a critical role in minimizing the use of chemicals and their environmental impacts, while optimizing beneficial results.

Management of Post-harvest Pests

Stored bamboos and finished products are susceptible to attack by boring insects, mainly shot-hole, powder-post and long-horned beetles. This is a serious problem in most bamboo-growing nations. Effective control measures are reported from many countries, and can be grouped as pre-harvest and post-harvest treatments, the latter involving prophylactic measures.

Pre-harvest treatment involves mainly fertilization and timing of harvesting. There are reports that correlate the damage level in felled culms to application of fertilizers in bamboo stands, culm age and season of felling, although not much experimental data are available to support this. It is generally believed that attack of borers on bamboos is highest when the harvesting is done during the full moon phase. Some studies have shown that the moisture level in culms increases with the waning of the moon and decreases with the waxing of the moon. This fortnightly rhythm in relative moisture content is the only lunar periodicity known to occur in the metabolism of the growing bamboo. However, it may be added that no relation has been found between the moisture content of freshly cut bamboo and its subsequent susceptibility to borer attack. Nevertheless, felling of bamboos based on the moon's phases is a traditional method practised to reduce borer incidence.



The correlation of the starch or total sugar content of the bamboo and beetle attacks is a point of much debate (Dhamodharan et al. 1986; Kumar et al. 1994). Some traditional methods of checking insect incidence by controlling starch content include felling of bamboo when the sugar content is low, felling of bamboo at full maturity, post-harvest transpiration of bamboo culms, and water soaking of bamboo to leach out soluble sugars and carbohydrates. Singh (1974) showed that starch content of bamboo is not related to the age of the culm, although flowered bamboo may show a reduction in the starch content. There are also many factors other than the starch content that may influence borer attack, and how these factors interact with the starch content needs to be studied in detail.

Prophylactic measures include treatment with cypermethrin 0.4% in diesel oil or endosulphan 0.75% in diesel oil against ghoon borer attack (Thapa et al. 1992); treatment with 1% sodium pentachlorophenate or sodium pentachlorophenate:boric acid:borax mixture (Kumar et al. 1985), etc. It is also suggested that prophylactic measures be adopted before the monsoon and treated stacks be stored under rain-proof shelters. Several preservative treatments have also been recommended against storage pest of bamboos. These have been discussed earlier in the section on Post-harvest Pests.



10. ENTOMOLOGICAL RESEARCH ON BAMBOO IN ASIA

Research Activities and Achievements

Bamboo is known as the poor man's timber and mostly grown in natural or less managed stands. Consequently, insect pest problems and studies on them have generally been ignored. The pioneer publications on pest problems on bamboos came from Japan at the end of the 19th Century, and subsequently from India and China during the early part of this century. These mostly noted the pest species and their damaging effects, but a few described the biology of insect pests in detail. Research activities on bamboo entomology have been mostly undertaken in China, India and Japan. Among 250 or so research papers on insect pests of bamboo in Asia published since the 1940s and abstracted by CAB, over 95% came from these three countries. In China, where serious pest problems have been reported in bamboo plantations, studies aiming to solve the problems received great attention (over 600 reports and papers related to bamboo entomology, but mostly in Chinese and published only in the country). In other countries, available information seems to be scattered. The poor response to a questionnaire, designed by the authors to collect existing data on bamboo pests, indicate that there are either no serious pest problems in these countries or they do not receive much attention.

Bamboo entomological research in Asia mainly covers the taxonomy and biology of insect pests and their natural enemies, and control measures.



Approximately 100 new species or genera of insects associated with bamboos have been identified and described by taxonomists in different Asian countries or regions. Entomologists have also prepared lists of bamboo insect pests in their own specific fields: Liao Huitang (1976) noted 23 species of bamboo aphids occurring in Taiwan-China; Yan Aojin et al. (1985) listed 137 species of scale insects feeding on bamboos in China; Ding Jinghua and Hu CL (1987) reported 31 species of bamboo leafhoppers; Xu Tiansen et al. (1993) listed 683 insect pest species on bamboos in China. The insect pests of bamboos and control measures in India were summarized by Singh and Bhandari (1988). Mathew and Nair (1990) and Mathew and Varma (1990) reported the insect pests in newly established bamboo plantations and those attacking post-harvest bamboos in Kerala, India.

Many of the research works on bamboo entomology report the results of investigations on the biology and control of major insect pests. The life history, biological characteristics and habits of about 100 insect species of both insect pests and their natural enemies have been described in detail in different countries, especially in China and Japan. Investigations on the ecological aspects of some insect pests, their relationships with host bamboos, natural enemies and other environmental conditions, have also been undertaken. Control and management measures against outbreaks of important pests on both standing and felled bamboos have been developed and are practised widely in some countries. Xu Tiansen (1992b) and Singh and Bhandari (1988) have reviewed the control measures practised in China and India, respectively.

The damage caused by boring insect pests to post-harvest bamboos is undoubtedly a serious problem



facing the bamboo industry in most of Asian countries, and the beetle species *Dinoderus* is responsible for much of the damage caused. Detailed investigations on the powder-post beetles — covering their occurrence, biology and control measures — have been undertaken and results well-documented in several countries, such as India, Japan, China and Indonesia. Some effective control measures have been developed and are in practice.

Major Existing Problems

Bamboo grows fast and, when managed well, its high productivity makes it a crop of substantial economic value, especially in rural areas of developing countries. There is also a rapidly expanding market abroad for bamboo products. Therefore, attempts to improve the productivity of bamboo stands through means such as plantation development and intensive silvicultural measures can be expected to increase in the immediate future. Consequently, it is also prudent to expect that the insect pest problems of standing bamboos will become more significant. However, insect pest problems in bamboo industry has received little attention and the pest status in most Asian countries is not fully known.

Information on the biology or effective control measures of some important insect pests are not available yet. An example is the seed bug, *Udonga montana*, which can damage all seeds during massive outbreaks. The situation is similar in the case of some sap-sucking insects, and particularly the shoot and culm borers found in many Asian countries. The damages, which appear to be more substantial in natural or less managed stands than in plantations, usually result in substantial reduction in both quality



and quantity of culms and edible shoots. The bamboo shoot weevil, *Otidognathus davidis*, is successfully controlled by injection of systemic insecticides in China, but there are no effective and simple measures for controlling outbreaks of most of the borers, such as the large bamboo shoot weevils (*Cyrtotrachelus spp.*), the shoot-boring noctuids, the bamboo hispine beetle (*Estigmena chinensis*) and the shoot-boring flies.

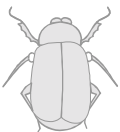
It is well recognized that the abundance, population dynamics and survival of insect pests are closely related to various environmental factors. However, there is an abject lack of knowledge on the ecological aspects of bamboo insect pests. The relationship and interaction among the insect pests and host food quality, natural enemies and physical factors are not known for most of the bamboo pests.

Another problem is that most of the control measures recommended for insect pests involve the application of chemical insecticides. Large-scale application of insecticides is repeatedly employed when bamboos are grown in plantations for commercial purposes, such as in China. The large amounts of chemical insecticides used create serious environmental problems, play havoc with natural pest control systems, and pose health hazards to humans and animals.

Recommendations for Future Research

The most urgent need is to extend available research achievements and to develop effective control measures against some of the important pests. For the long-term, studies on ecological aspects and integrated pest management should be encouraged.

It is not difficult to recognize the importance of extension programs. Research activities on bamboo



entomology in different Asian countries vary much. Besides, detailed studies and notable research achievements are restricted to a few countries. These results are valuable and applicable in many countries as some insect pests are widespread in Asia. Extension programs can be designed:

- **1** to extend available knowledge on the identification, biology and control of important insect pests on bamboos; and thus,
- **2** to help identify major insect problems, their current and potential threats to the bamboo industry, and
- **3** to solve some of the major existing problems in many countries, where little relevant studies have been conducted.

It is also urgent that effective and practical control measures be developed against some of the major insect pests, especially those that bore bamboo shoots and culms. In small-scale stands, they are traditionally controlled through direct measures such as hand-picking or destroying the damaged plants, but these measures are not practical in plantations. Silvicultural and biological approaches are the most promising in the control of these borers and need to be investigated.

Much attention has been given to pest problems related to post-harvest bamboos, mainly because of economic reasons. The present pest management approaches are based on chemical treatment, and the use of less persistent chemicals and alternatives is an area of research that needs to be developed. Also, traditional knowledge, wherever applicable, may be tested and popularized.

The long-term strategy for releasing pest pressure on the bamboo industry is to develop IPM programs.



But studies in some basic areas need to be undertaken first. The impact of insects on productivity and economics requires to be evaluated. The effects of various environmental conditions on insects — including silvicultural measures, host food quality, natural enemies and physical factors, as well as their interactions — need to be investigated and fully understood. Such data are essential to the development of IPM strategies. In undertaking these extension and research programs, international and regional cooperation are essential.



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PICTURE/ILLUSTRATION CREDITS

Mr Wang Haojie

Mr Subash Kuriakose

Mr Xu Tiansen

Mr Xiang Zhang

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REFERENCES

- Agarwala, B.K. 1981. Note on some aphids affecting economically important plants in Sikkim. *Indian Journal of Agricultural Sciences*, 51(9), 690-692.
- Anonymous. 1960. Population dynamics of bamboo locust and integrated control measures. *Entomological Knowledge*, 6(2), 65-67.
- Anonymous. 1976. Study on *Loudonta dispar*. *Entomological Knowledge*, 6(3), 88-89.
- Anonymous. 1977. A preliminary study on bamboo gall-making chalcids. *Scientia Silvae Sinicae*, 13(4), 56-62.
- Anonymous. 1978. Studies on biology and control of *Allobremeria plurilineata*. *Journal of Hunan Forestry Science and Technology*, 2, 31-32.
- Anonymous. 1979. Culm cavity injection of systemics for control bamboo leaf-rollers. *Forest Science and Technology*, 3, 19-20.
- Beeson, C.F.C. 1941. The ecology and control forest insects of India and neighbouring countries. Vasant Press, Dehra Dun, India. 30, pp. 113-145.
- Bhasin, G.D.; Roonwal, M.L.; Singh, B. 1958. A list of insect pests of forest plants in India and the adjacent countries. Part 3: list of insect pests of plant genera 'A' (Appendix only), 'B' (Baccavrea to Bazus) and 'C' (in part) (Cadaba to citrus). *Indian Forestry Bulletin* (N.S.), 171(2), (Ent.), 1-126.
- Bhatia, D.R.; Singh, P. 1962. Beetles as predators on the desert locusts. *Indian Journal of Entomology*, 24(1), 103-108.



- Browne, F.G. 1968. Pest and diseases of forest plantation trees. Clarendon Press, Oxford, UK. 1330 pp.
- Cai Rongquan. 1986. Identification of bamboo puss moths. *Forest Pests and Diseases*, 1, 38-42.
- Cai Xiaoyang. 1988. Study on bionomics of the bamboo gall-making midge, *Planetella conesta*. *Acta Entomologica Sinica*, 31(4), 439-441.
- Cao Zhifang; Xia, L.J. 1987. On integrated measures against bamboo leaf rollers. *Journal of Jaingsu Forestry Science and Technology*, 2, 31-33.
- Chang Lefeng et al. 1979. A study on the biology and control of the powder-post beetle *Dinoderus japonicus* Lesne. *Acta Entomologica Sinica*, 22(2), 127-132.
- Chang Yuzhen. 1981. The morphology, damage and control of the bamboo mirid bug, *Mecistoscolis scitetooides* (Hemiptera: Miridae). *Bulletin of Plant Protection*, 23 (1), 15-23.
- Chang Yuzhen. 1986. Insect pests of bamboos in Taiwan. In Higuchi, T. ed., 1986. Bamboo production and utilization. Proceedings of the Congress Group 5.04, production and utilization of bamboo and related species, XVIII IUFRO World Congress Ljubljana, Yugoslavia, 7-21 September 1986. Kyoto University, Kyoto, Japan. pp. 246-252.
- Chang Yuzhen; Xue XQ. 1994. The distribution, damage and control of major forest insect pests in Taiwan. *Journal of Fujian Forestry Science and Technology*, 21(1), 44-49.
- Chatterjee, P. N.; Misra, M.P. 1974. Natural insect enemy-plant host complex of forest insect pest of



Indian region. Indian Forest Bulletin, 265 (N.S.) (Ent.), 1-233.

■ Chatterjee, P.N.; Sebastian, V.O. 1964. Notes on the outbreak of sap sucker *Oregma bambusae* Buckt in New Forest and measures taken to control them. Indian Forester, 90(1), 30-31.

■ Chaudhry, G.U. 1955. Notes on a collection of termites from Pakistan. Pakistan Journal of Forestry, 51(1): 40-43.

■ Chen Darong. 1983. A preliminary study on the bamboo tussock moth, *Pantana pluto* (Leech). Journal of Guangdong Forestry Science and Technology, 5, 24-26.

■ Chen, H.T. 1928. Notes on bamboo borer, *Cyrtotrachelus longimanus*. Lingnan Science Journal, 6, 353-366.

■ Chen Jiwen; Yang MZ; Liang DR; Cai YN. 1984. Morphological, pathological and toxicological determination of the granuloses virus in *Parasa bicolor*. Forest Science and Technology, 9, 24-26.

■ Chen Liang. 1983. Preliminary study on the lyctid beetle, *Lyctus brunneus*. Scientia Silvae Sinicae, (Mem.), 126-128.

■ Chen Mingfa; Cheng CL. 1992. Studies on the bionomics of *Oligonychus uruma* Ehara on bamboo. Chinese Journal of Entomology, 12(1), 21-29.

■ Chen Wenjie; Jiang YQ; Ding JS. 1988. Study on control of *Hieroglyphus tonkinensis*. Journal of Fujian Forest Science and Technology, 2, 19-24.

■ Chen Wenjie; Jiang, YQ; Ding JS. 1989. Study on biology of *Hieroglyphus tonkinensis*. Forest Insects and Diseases, 2, 14-25.



- Chen Yangchun. 1982. Preliminary study on *Pantana phyllostachysae*. *Scientia Silvae Sinicae* 18(3), 343-346.
- Chen Yijin. 1982. Studies on the bamboo shoot noctuid (*Oligia apameoides* Drt.). *Scientia Silvae Sinicae*, 18(2), 151-159.
- Chen Zhenyao. 1989a. Notes on Hemipterian insects feeding on bamboos in Guangdong, China. *Journal of Bamboo Research*, 8(3), 58-60.
- Chen Zhenyao. 1989b. Study on bionomics of the bamboo coreid bug, *Notobitus meleagris*. *Entomological knowledge*, 26(4), 226-228.
- Chien Tingyu. 1982. Studies on *Dorysthenes (Baladeva) walkeri* Waterhouse. *Acta Entomologica Sinica*, 25(1), 31-34.
- Choldumrongkul, S. 1994. Insect pests of bamboo shoot in Thailand. *In Bamboo in Asia and the Pacific. Proceedings of the 4th International Bamboo Workshop, Chiangmai, Thailand, 27-30 November 1991. International Development Research Centre, Ottawa, Canada; Forestry Research Support Programme for Asia and the Pacific, Bangkok, Thailand. pp. 331-335.*
- Cui Yunji. 1989. Notes on leaf mites on bamboos in China. *Entomological Knowledge*, 5, 303-304.
- Dayan, MP. 1990. Survey, identification and pathogenicity of pests and diseases of bamboo in the Philippines. Paper presented at the First National Bamboo Symposium Workshop, 27 Feb-1 March 1989. *Sylvatrop*, 13(1-2): 61-67.
- Dayun, W.; Shen Shaojin. 1987. *Bamboos of China*. Christopher, London, UK. 167 pp.
- Dhamodharan, T.K.; Mathew, G.; Gnanaharan, R.; Nair, K.S.S. 1986. Relationship between starch content



and susceptibility to insect borer in the bamboo reed, *Ochlandra travancorica*. Entomon, 11(4), 215-218.

■ Ding Daomo; Zhang Shimei. 1956. Notes on incidence and control measures of bamboo locusts in Jiangxi Province. Entomological Knowledge, 2(5), 217-220.

■ Ding Jinghua; Hu CL. 1987. A preliminary list of plant hoppers attacking bamboo in China with an illustrated key to the genera and species (Homoptera: Delphacidae). Journal of Bamboo Research, 6(4), 70-84.

■ Fang Li; Yuan ZT. 1992. The bionomics and control of bamboo noctuid defoliator, *Eustotia* sp. Journal of Shandong Forestry Science and Technology, 3, 44-46.

■ Fei Xueqing; Xu Tiansen; Li WX. 1990. Changes of nutrients in bamboo twigs damaged by *Hippotiscus dorsalis* (Stal.). Forest Research, 3(5), 509-512.

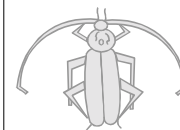
■ Feng Zongxuan et al. 1987. Brief note on biology of *Kuwanaspis howarai* (Cooley). Forest Pests and Diseases, 3, 1-2.

■ Fletcher, T.B. 1920. Life-histories of Indian insects: Microlepidoptera. Mem. Dept. Agri. Indian (Ent. Ser.), 6, 1-12.

■ Gan Zhongnan. 1984. A Preliminary study on the bamboo cicada *Platylocma pieli* Kato. Forest Pests and Diseases, 4, 5-7.

■ Gao Zhaowei. 1979. Note on the occurrence of the Japanese bamboo lygaeid bug, *Pirkimerus japonicus*, on *Phyllostachys pubescens*. Journal of Zhejiang Forestry Science and Technology, 2, 39-40.

■ Gao Zhaowei. 1980. Studies on the morphology, life-cycle and control of *Pirkimerus japonicus*. Journal of Zhejiang Forestry Science and Technology, 4, 5-7.



- Garthweite, P.E. 1938. Entomological research. Rep. Sil. Ent. Burma 1936-1937. pp. 93-103.
- Guo Peide. 1989. Study on trapping against *Ceracris kiangsu*. Journal of Hunan Forest Science and Technology, 1, 45-46.
- Gupta, Y.N. 1978. On a collection of tetranychoid mites from India with a description of a new *Eotetranychus*. Indian Journal of Acarology, 3(2), 87-91.
- Hamon, A.B. 1980. Bamboo pit scale, *Asterolecanium bambusae* (Boisduval) (Homoptera: Coccidae: Asterolecaniidae). Entomology Circular No. 220. Division of Plant Industry, Department of Agriculture and Consumer Service, Florida, USA.
- Hu Heyuan; Min ZJ. 1988. Study on control of *Nesticoccus sinensis* using dimethoate. Journal of Jiangsu Forestry Science and Technology, 1(32), 19.
- Huang Ertian. 1984. Studies on control of bamboo leaf-rollers by light-trapping and artificial releasing *Trichogramma dendrolimi*. Natural Enemies of Insects, 6(2), 118-120.
- Huang Jinshui; Lin QY. 1992. Notes on *Matapa aria* Moore. In Xiao Gangrou, ed., Forest Pests of China. China Forestry Publishing House, China. pp. 1140-1141.
- Huang Rizong. 1963. Preliminary study on the bamboo red long-horned beetle, *Purpuricenus temminckii* G.-M. Entomological Knowledge, 7(1), 26-27.
- Huang Zenghe; Wu JF; Dun CF. 1982. Biology of *Hieroglyphus tonkinensis* and control experiments. Journal of Bamboo Research, 1(2), 62-71.
- Hutson, J.C. 1933. Report on the work of entomological division. Rep. Ceylon Dept. Agri., 1-23.



- Jin Changle; Chen YD; Zhang ZZ. 1980. Releasing *Trichogramma dendrolimi* against outbreak of bamboo leaf-roller. Journal of Zhejiang Forestry Science and Technology, 2, 29-33.
- Kaneko, T. 1959. Egg-laying of *Oligia (Procus) vulgaris* and its control. Journal of Japanese Forest Science, 414, 148-148.
- Kim, K.C.; Lee, T.S. 1986. Studies on the host plant, bionomics, and damage of bamboo leaf rollers in Chonnam Province area. Korean Journal of Plant Protection, 25(2), 85-92.
- Kumar, S.; Kalra, K.K.; Dobriyal, P.B. 1985. Protection of pulp-bamboo in outside storage, Journal of Timber Development Association (India), 31(4), 5-12.
- Kumar S.; Shukla, K.S.; Indra Dev; Dobriyal, P.B. 1994. Bamboo preservation techniques: a revision. INBAR Technical Report No.3. International Network for Bamboo and Rattan, New Delhi, India; Indian Council of Forestry Research and Education, Dehra Dun, India. pp. 59.
- Lan Linfu. 1980. Cavity injection of systemics: an effective control method against bamboo leaf-rollers. Journal of Zhejiang Forestry Science and Technology, 1, 15.
- Lan Shiwen et al. 1993. Study on biology and control of *Lethe europa* feeding on bamboo. Forest Research, 6(1), 112-116.
- Liao GL; Shen GP. 1981. Preliminary study on *Leptispa godwini* Baly. Entomological Knowledge, 18(4), 167-179.
- Liao Huitang. 1976. Bamboo aphids of Taiwan. Quarterly Journal of the Taiwan Museum, 29(3-4), 499-586.



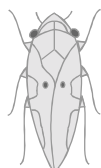
- Liao Xueen. 1984. Preliminary study on the bamboo grey tent caterpillar moth. *Forest Pests and Diseases*, 2, 21-22.
- Liu Jinying. 1956. Notes on bostrychid borers on bamboos. *Entomological Knowledge* 2(1), 221-224.
- Liu Nanxing; Zhang ZY; Zheng LS. 1988. A preliminary test on control of bamboo shoot weevils, by using nematodes. *Journal of Guangdong Forestry Science and Technology*, 4, 32-33.
- Liu Nanxing; Zhang ZY; Zheng LS. 1989. Study on the entomopathogenic nematodes for biological control of bamboo shoot weevil, *Cyrtotrachelus longimanus* Fab. (Coleoptera: Curculionidae). *Natural Enemies of Insects*, 11(1), 44-50.
- Liu Ruilan. 1988. Control of *Algedonia coclesalis* with *Trichogramma dendrolimi* Matsumara. *Journal of Zhejiang Forestry Science and Technology*, 8(2), 40-42.
- Liu Yuan; Xu FM 1982. Preliminary study on life cycle of *Dinoderus minutus* Fab. *Journal of Sichuan Forestry Science and Technology*, 12, 26-28.
- Liu Yuan; Xu FM. 1985. Preservative measures against infestation by insect pests and fungi. *Bamboo Research* (suppl.), 55-59.
- Liu YC; Chang Yuzhen. 1976. Population dynamics of *Astegopteryx bambusifoliae* Takahashi (Homoptera: Aphidae). *Journal of Agriculture and Forestry*, 25, 71-84.
- Liu Yongzhen. 1978. Preliminary study on the large bamboo shoot weevil. *Acta Entomologica Sinica*, 21(2), 204.
- Lu Ruoqing; Xu Tiansen. 1988. Preliminary study on *Glyphipterix semiflavana* mining bamboo leaves. *Forest Pest Disease*, 4, 13-14.



- Lu Ruoqing; Xu Tiansen. 1992. Studies on the bionomics of *Aphrophora horizontalis* Kato and its control. *Forest Research*, 5(6): 687-692.
- Ma Junchao. 1934. Notes on bamboo shoot-boring noctuids in Zhejiang. *Plant Insects and Diseases*, 2(36), 709.
- Main, J.F. 1912. Campaigns against rice grasshopper. *Agricultural Journal of India*, 7(3), 246-256.
- Mathew, G.; Sudheendrakumar. 1992. Outbreak of *Udonga montana* (Heteroptera: Pentatomidae) on bamboo in natural forests and adjoining plantation in Wynad. *Bamboo Information Centre- India Bulletin*, 2(2), 17-18.
- Mathew, G.; Nair, K.S.S. 1990. Storage pests of bamboo in Kerala. *In* Ramanuja Rao, I.V.; Gnanaharan, R.; Sastry, C.B. ed., *Bamboo: current research*. Proceedings of the International Bamboo Workshop, Cochin, India, 14-18 November 1988. Kerala Forest Research Institute, Kerala, India; International Development Research Centre, Ottawa, Canada. pp. 212-214.
- Mathew, G.; Varma, R.V. 1990. Occurrence and pest status of some insects attacking bamboos in newly established plantations in Kerala. *In* Ramanuja Rao, I.V.; Gnanaharan, R.; Sastry, C.B. ed., *Bamboo: current research*. Proceedings of the International Bamboo Workshop, Cochin, India, 14-18 November 1988. Kerala Forest Research Institute, Kerala, India; International Development Research Centre, Ottawa, Canada. pp. 195-198.
- Mathur, R.N. 1943. Bamboo defoliators. *Indian Journal of Entomology*, 5(1), 117-128.



- Meshram, P.B.; Joshi, K.C.; Namdeo, P.K. 1989. White grub, *Holotrichia* sp., a pest of bamboos in Orissa. Paper submitted at Seminar Silviculture and Management of Bamboos. Institute of Deciduous Forests, Jabalpur, India. pp. 93-94.
- Mo Jianchun et al. 1992. Studies on chemical control of bamboo eurytomid and torymid. *Forest Science and Technology*, 9, 12-14.
- Mori, H.; Hideo, A. 1979. Insect damage to bamboo materials and its prevention. *Science for Conservation*, 18, 41-55.
- Nair, K.S.S.; Mathew, G.; Varma, R.V.; Gnanaharan, R. 1983. Preliminary investigations on the biology and control of beetles damaging stored reed. KFRRI Research Report 19. Kerala Forest Research Institute, Kerala, India. 35 pp.
- Nakahara, J.; Kobayashi, F. 1963. Taxonomy and biology of bamboo leaf rollers (Pyraustinae) *Bulletin of Forestry Experiment Station, Meguro, Tokyo*, No. 151, 45-52.
- Pruthi, H.S. 1939. The bionomics, life history and control of the grasshopper, *Poecilocerus pictus*, a new pest of cultivated crops in north India. *Indian Journal of Agricultural Science*, 9, 629-642.
- Qui Zilin; Hu YL. 1982. Study on *Pirkimerus japonicus*. *Forest Science and Technology*, 4, 30-32.
- Rao, Y.R. 1960. The desert locust in India. I.C.A. Entom. Monograph No.21, 721 pp.
- Rao, Y.R.; Cherian, M.C. 1940. Control of the rice grasshopper. *Ind. For. Farm.*, 1(10), 495-498.
- Roonwal, M.L. 1970. Termites of oriental regions. *In* Krishna, K.; Weesner, F.M., ed., *Biology of Termites*. Vol. II. pp. 315-391. Academic Press, New York., USA.



- Roonwal, M.L. 1977. Field ecology and biology of the bamboo beetle, *Estigmene chinensis* Hope (Coleoptera: Chrysomelidae) in the western sub-Himalayas. *Journal of Entomological Research*, 1(2), 168-175.
- Roonwal, M.L. 1979. Termite life and termite control in tropical South Asia: XI. pp. 1-177.
- Saito, Y.; Ueno, J. 1979. Life history studies on *Schizotetranychus celarius* (Banks) and *Aponychus* mite species. *Applied Entomology and Zoology*, 14(4), 445-452.
- Sandhu, G.S. 1974. Studies on insects infesting wooden and bamboo household articles and their control. *Journal of Research, Punjab Agricultural University, Punjab, India*, 12(2), 152-155.
- Sen-Sarma, P.K.; Mathur, R.N. 1957. Further record of occurrence of *Cryptotermes dudleyi* (Banks) in India (Isoptera: Kalotermitidae). *Current Science*, 26 (12), p. 399.
- Sen-Sarma, P.K.; Thakur, M.L.; Mishra, S.C.; Gupta, B.K. 1975. Studies on wood-destroying termites in relation to natural termite resistance of wood. Final Technical Report, PL 480 Project A-7-58 (1968-73), Dehra Dun, India.
- Shen Jizeng. 1964. Preliminary study on *Ceracris nigricornis*. *Scientia Silvae Sinicae*, 9(3), 272-274.
- Shen Jizeng. 1965. Study on biology of *Ceracris kiangsu*. *Entomological Knowledge*, 10(2), 84.
- Shi Jimao; Yu Huaxing; Yang SD. 1992. Studies on the bionomics of three species of bamboo spider mites and their control techniques. *Journal of Zhejiang Forestry Science and Technology*, 12(2), 11-14.



- Shi Quantai. 1980. Country report on China. *In* Lessard, G; Chouinard, A. ed., *Bamboo research in Asia. Proceedings of a workshop held in Singapore, 28-30 May 1980.* International Development Research Centre, Ottawa, Canada. pp. 57-62.
- Singh, B.; Tewari, M.C. 1979. Studies on the treatment of bamboos by steeping, open tank and pressure. *Journal of Indian Academy of Wood Science*, 10(2), 68-71.
- Singh, B.; Tewari, M.C. 1981a. Studies on the treatment of green bamboo by steeping and sap-displacement method. *Journal of Indian Academy of Wood Science*, 2(1), 21-27.
- Singh, B.; Tewari M.C. 1981b. Studies on the treatment of green bamboo by different diffusion processes- Part II. *Journal of Timber Development Association (India)*, 27 (2), 38-46.
- Singh, P. 1990. Current status of pests of bamboos in India. *In* Ramanuja Rao, I.V.; Gnanaharan, R; Sastry, C.B., ed., *Bamboos: current research.* Proceedings of the International Bamboo Workshop, Cochin, India, 14-18 November 1988. Kerala Forest Research Institute, Kerala, India; International Development Research Centre, Ottawa, Canada. pp. 190-194.
- Singh, P.; Bhandhari, R.S. 1988. Insect pest of bamboos and their control. *Indian Forester*, 114(10), 670-683.
- Singh, S. 1974. The incidence of borer attack on felled bamboos - some investigations and observations. *Indian Pulp and Paper*, 29(2), 7-14.
- Stapleton, C.M.A. 1985. Noctuid shoot borers in *Dendrocalamus* and *Bambusa* species. *Nepal Forestry Technical Information Bulletin*, 11, 26-31.



- Sulthoni, A. 1990. A simple and cheap method of bamboo preservation. *In* Ramanuja Rao, I.V.; Gnanaharan, R; Sastry, C.B., ed., Bamboos: current research. Proceedings of the International Bamboo Workshop, Cochin, India, 14-18 November 1988. Kerala Forest Research Institute, Kerala, India; International Development Research Centre, Ottawa, Canada. pp. 209-211.
- Takahashi, F.; Mizuta, K. 1971. Life cycle of a eurytomid wasp, *Aiolomorphus rhopaloides* and three species of wasp parasitic on it. *Japanese Journal of Applied Entomology and Zoology*, 15(1), 36-43.
- Tan Zhengyi. 1984. The bionomics of *Dinoderus minutus*. *Bamboo Research*, 1, 83-84.
- Tao Dexin; Zhang AL. 1989. The biology and control of *Hippotiscus dorsalis*. *Journal of Bamboo Research*, 4, 65-73.
- Thakur, M.L. 1988a. Current status of termites as pest of bamboos and their control. *Indian Forester*, 114(10), 720-725.
- Thakur, M.L. 1988b. Pest problems in forest nurseries and their management. *In* Gupta V.K.; Sharma, N.K., ed., *Tree Protection*. Proceedings of the National Seminar on Tree Protection. Y.S. Parmar University of Horticulture and Forestry, Solan, Himachal Pradesh, India. pp. 332-341.
- Thapa, R.S.; Singh, P.; Bhandari, R.S. 1992. Prophylactic efficacy of various insecticides for the protection of bamboos in storage against ghoon borers, *Dinoderus* sp. *Journal of Indian Academy of Wood Science*, 23(1).
- Togashi, I. 1974. Studies on the pests of *Phyllostachys heterocyclus* (Carr): I. Life history of *Cosmopterix*



phyllostachysae Kuroto (Cosmopterigidae) and its natural enemies. Applied Entomology, 9(2), 101-103.

■ Varma, R.V.; Mathew, G.; Mohandas, K.; Gnanaharan, R.; Nair, K.S.S. 1988. Laboratory evaluation of insecticides for the control of the bamboo borers, *Dinoderus minutus* and *D. ocellaris* (Coleoptera: Bostrychidae). Material und Organismen, 23(4), 281-288.

■ Wang Haojie; Wu ZY; Lin CC; Xu Tiansen. 1995. Host range of the bamboo shoot weevil, *Otidognathus davidis* (Coleoptera: Curculionidae). Forest Research, 8(3), 309-313.

■ Wang Haojie; Xu Tiansen et al. 1993. Studies on the effect of methamidophos on *Otidognathus davidis*. Forest Research, 6(3), 337-340.

■ Wang Haojie; Xu Tiansen; Lin CC; Liu RP. 1996a. Studies on the biological characteristics of two chalcid-flies infesting bamboo. Forest Research 9(1), 52-57.

■ Wang Haojie; Xu Tiansen; Lin CC; Liu RP. 1996b. Studies on interspecific relationships of two chalcid-flies on bamboo. Forest Research, 9(3). (in press).

■ Wang Haojie; Xu Tiansen; Wang GH. 1994. Development of insecticide-injection technique against *Otidognathus davidis* Fair. on bamboo. Forest Research, 7(mem), 103-108.

■ Wang Maozhi. 1989. Preliminary study on *Eutomostethus nigrinus* Xiao. Forest Science and Technology, 1, 24-27.

■ Wang Shufen; Liu SB; Li BS. 1990. Epidemic interval and forecasting of *Ceracris kiangsu*. Journal of Central-South Forestry College, 10(1), 1-6.

■ Wang Ziqing; Zhang XJ. 1987. Identification of important species in the genus *Bambusaspis*. Forest Pests and Diseases, 3, 36-41.



- Wei Houjian. 1984. Investigation on the Chinese bamboo tussock moth. *Journal of Bamboo Research*, 3(1), 64-67.
- Wei Shanqun. 1996. Preliminary study on control of bamboo skeletonizers. *Scientia Silvae Sinicae*, 3, 230-231.
- Wu Jianfen. 1984. Study on the granulosis virus disease infesting *Brtona funeralis*. *Journal of Central-South Forestry College*, 4(2), 156-157.
- Wu Jianfen; Huang Zenghe. 1986. A Preliminary study on the bostrychid, *Dinoderus minutus* Fab. *Journal of Bamboo Research*, 5(1), 112-119.
- Wu Jianfen; Huang Zenghe; Chen SH. 1984. A preliminary study of eucleid *Parasa bicolor* (Walker). *Journal of Bamboo Research*, 3(2), 120-124.
- Wu Jianfen; Wang Shufen. 1980. A preliminary study of the bamboo zygaenid, *Brtona funeralis* Butler. *Scientia Silvae Sinicae*, 16(suppl), 77-83.
- Wu Shijun. 1981. A preliminary study on the bamboo scale *Unachionaspis bambusae*. *Entomological Knowledge*, 4, 163-164.
- Wu Shijun. 1992. Notes on new species of *Bambusaspis*. *Acta Entomologica Sinica*, 35(1), 75-77.
- Wu Shixiong. 1979. Biology of large bamboo shoot weevils. *Entomological Knowledge*, 16(6), 256.
- Xi Gengsi; Zheng ZM. 1989. A preliminary study on *Parnara ganga* Evans injuring bamboos. *Journal of Bamboo Research*, 8(1), 52-56.
- Xiao Caiyu. 1963. Notes on coreid bug species feeding on bamboo in China. *Acta Entomologica Sinica*, 12(4), 506-510.
- Xiao Gangrou. 1954. Population dynamics and control measures of bamboo. *New Science*, 2, 38-43.



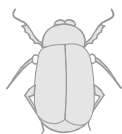
- Xiao Gangrou. 1959. Review on researches and control measures of bamboo locusts. *In* Entomological Paper Collection. Scientific Press. pp. 38-45.
- Xie Guolin. 1983. Studies on *Nesticoccus sinensis*. *Acta Entomologica Sinica*, 26(3), 268-277.
- Xie Qinmei, 1983. Isolation and observation of nuclear polyhedrosis virus of *Pantana phyllostachysae* Chao. *Forest Science and Technology*, 2, 26-27.
- Xing Longping. 1980. The morphology, occurring and control of *Pantana phyllostachysae*. *Forest Science and Technology*, 7, 26-28.
- Xu Fuyuan. 1981. Study on *Parasa bicolor*. *Journal of Jiangsu Forestry Science and Technology*, 4, 9-12.
- Xu Ji. 1983. Preliminary studies on scale insects on bamboos. *Journal of Shanxi Forestry Science and Technology*, 4, 47-56.
- Xu Tiansen. 1964. Biology and control of the bamboo shoot weevil. *Journal of Zhejiang Agricultural Science*, 2, 75-78.
- Xu Tiansen. 1983. Integrated measures for control of insect pests on post-harvest bamboo. *Journal of Subtropical Forestry Science and Technology*, 3, 50-53.
- Xu Tiansen. 1984a. Study on *Ceracris kiangsu* in Zhejiang. *Forest Insects and Diseases*, 4, 20-22.
- Xu Tiansen. 1984b. About the grey tent caterpillar moth, *Cosmotriche* sp. *Journal of Subtropical Forestry Science and Technology*, 4, 44-45.
- Xu Tiansen. 1985a. Study on bionomics of *Eumorphobotys obscuralis*. *Journal of Subtropical Forestry Science and Technology*, 3, 55-56.
- Xu Tiansen. 1985b. Study on the biological characteristics of Chinese tussock moth, *Pantana sinica* Moore. *Journal of Bamboo Research*, 4(1), 66-76.



- Xu Tiansen. 1987. Notes on three bug species feeding on bamboo. *Forest Pests and Diseases*, 4, 33-35.
- Xu Tiansen. 1989. A preliminary study on *Crocidophora evenoralis* on bamboos. *Forest Pests and Diseases*, 1, 21-23.
- Xu Tiansen. 1992a. Notes on *Circobotys aurealis* and *Demobotys pervulgaris*. In Xiao Gangrou, ed., *Forest insects of China*. China Forestry Publishing House, China. pp. 857-863.
- Xu Tiansen. 1992b. Reviews of studies on control measures against bamboo insect pests in China. *Journal of Shanxi Forestry Science and Technology*, 2, 40-43.
- Xu Tiansen; Ge Zhenghua. 1966. Survey on death of new shoots in relation with the bamboo shoot fly *Pegomyia phyllostachys* Fan. *Scientia Silvae Sinicae*, 11(3), 188-195.
- Xu Tiansen; Lu Ruoqing. 1987a. Study on the biological characteristics of *Phlaeoba angustidorsis*. *Journal of Bamboo Research*, 6(2), 69-76.
- Xu Tiansen; Lu Ruoqing. 1987b. A study on the *Loudonta dispar* (Kiriakoff). *Journal of Subtropical Forestry Science and Technology*, 3, 194-203.
- Xu Tiansen; Lu Ruoqing. 1988. A study on *Chyliza bambusae*. *Forest Research*, 1(3), 278-284.
- Xu Tiansen; Lu Ruoqing. 1990. Studies on *Besaia goddrica* (Lepidoptera, Notodontidae). *Forest Research*, 3(6), 568-573.
- Xu Tiansen; Lu Ruoqing. 1991. A preliminary research on the biological habits and characteristics of *Neope muirheadi* Felder. *Journal of Bamboo Research*, 10(2), 47-53.



- Xu Tiansen; Lu Ruoqing; Lin SS. 1988. A study on *Hippotiscus dorsalis* (Stal), I. Biology. Forest Research, 1(6), 634-640.
- Xu Tiansen; Lin SS; Lu Ruoqing. 1989. A study on *Hippotiscus dorsalis*, II. Enemies and chemical control. Forest Research, 2(2), 119-123.
- Xu Tiansen; Sun Yongchun. 1962. Studies on control measures against bamboo shoot-boring noctuids. Scientia Silvae Sinicae, 4, 284-291.
- Xu Tiansen; Sun Yongchun. 1978. Bionomics and control of *Algedonia coclesalis*. Scientia Silvae Sinicae, 1, 49-55.
- Xu Tiansen; Wang HJ; Lu Ruoqing. 1993. The revised list of insect pests on bamboos in China. Zhejiang Forest Pests, 4, 4-34.
- Xu Tiansen; Zhao Jingnian. 1974. Studies on bamboo leaf-rollers and their natural enemies. Journal of Subtropical Forestry Science and Technology, 2, 28-30.
- Xu Tiansen; Zhao Jingnian. 1976a. Studies on use of *Trichogramma dendrolimi* against bamboo leaf-rollers. Journal of Subtropical Forestry Science and Technology (1-2), 31-55.
- Xu Tiansen. Zhao Jingnian. 1976b. Notes on bamboo leaf-rollers in Zhejiang Province. Journal of Subtropical Forestry Science and Technology (3), 20-23.
- Yamamura, N. 1987. Biparental defence in a sub-social spider mite. Trends in Ecology and Evolution, 2(9).
- Yan Aojin. 1985. A preliminary list of scale insects on bamboos. Journal of Bamboo Research (suppl), 71-82.



- Yan Aojin. 1992. Biology of the bamboo pit scale *Bambusaspis hemisphaerica* (Kuwana). In Xiao Gangruo, ed., Forest insects on China. China Forestry Publishing House, China. pp. 293-294.
- Yan Aojin; Xie GL; Feng ZX. 1985. Studies on the bamboo scale *Kuwanaspis pseudoleucaspis* (Kuwana). Journal of Nanjing Institute of Forestry, 4, 41-50.
- Yang Guarong. 1991. Integrated control of bamboo forest insect pests. Project review report of Zhejiang Station for Forest Protection.
- Yang Jikun; Wang XL. 1988. A new species of *Chyliza* (Psilidae, Diptera) from China, injuring bamboo roots. Forest Research, 1(3), 275-277.
- Yang Mingzhen et al. 1984. A preliminary study on the bamboo scale *Kuwanaspis pseudoleucaspis*. Forest Pests and Diseases, 1, 9-11.
- Yao Kang; Hu WZ; Pan XD. 1986. Study on control of bamboo bostrychid borers through heating by infrared techniques. Entomological Knowledge, 23(2), 64-66.
- Yu Huaxing; Shi Jimao. 1991. Study on *Schizotetranychus nanjingensis*. Journal of Bamboo Research, 10(2), 61-67.
- Zeng Lin. 1981. Bionomics and control of *Notobitus montanus*. Entomological Knowledge, 18(1), 23-24.
- Zhan Zhongcai. 1983. Studies on bionomics and control of the Chinese powder post beetle *Lyctus sinensis* Lesne. Forest Pests and Diseases, 3, 25-28.
- Zhan Zhongcai. 1986. Preliminary study on *Ceresium sinicum*. Forest Pests and Diseases, 4, 1-2.
- Zhang Guoxian et al. 1992. A study on *Eriococcus rugosus*. Forest Research, 5(4), 429-435.



- Zhang Xiankai; Zuo YX. 1988. A preliminary study on *Niphona furcata* (Bates). *Entomological Knowledge*, 25(4), 218-220.
- Zhang Zongfu. 1984. A preliminary study on heteroecious habit of *Sagra femorata purpurea* Lichtenstein, *Bamboo Research*, 2, 34-37.
- Zheng Guohua et al. 1994. Damage and control of *Oligia vulgaris*. *Forest Research*, 7(1), 111-115.
- Zheng Hanyi. 1962. Study on control of shoot-boring noctuids. *Journal of Jiangsu Agricultural Science*, 4, 84-89.
- Zheng Jianjia; Yang GR; Jiang P. 1992. Field experiment on control of shoot-boring noctuids. *Journal of Bamboo Research*, 11(3), 32-35.
- Zheng Leyi; Wang ZF. 1987. New records on lygaeid bugs on bamboos in China. *Journal of Bamboo Research*, 6(4), 65-69.
- Zhou Fanchun. 1985. Bamboo preservation against infestation of boring pests and fungi. *Bamboo Research (suppl.)*, 60-67.
- Zhou Huiming et al. 1987. Studies of reasonable application of insecticide to control and cure the bamboo and bamboo wares damaged by *Dinoderus minutus*. *Journal of Nanjing Forestry University*, 4, 48-51.



GLOSSARY

Aestivation: Dormancy during hot and drought periods.

Cuckoo-spit: Also called **frog spit**. A frothy secretion made on plants by the young of certain insects, such as the frog hoppers, to function as a protective covering.

Diapause: A state of arrested development, especially in immature insects, often occurring seasonally or when environmental conditions are unfavourable.

Elytra: Thickened or horny front wing.

Epithelium: A cellular tissue covering the body surface.

Gall: An abnormal growth of plant tissues caused by the stimulus of an animal or another plant.

Grub: Larva of beetles.

Heteroecism: The development of different stages of a (pest) species on different host plants.

Inquilinous: A condition in which an animal lives in the nest of another species.

Instar: A young insect in one of its growth periods between moults.

Light-trapping: Collecting insects at night using traps fitted with light

Mandibles: A paired mouth appendage, usually a set of grinding jaws found in chewing insects.

Nematode: Any unsegmented worm (roundworm).

Nymph: The second stage of an insect undergoing incomplete metamorphosis (egg-nymph-adult



instead of the egg-larva-pupa-adult in complete metamorphosis). Most nymphs resemble their parents, unlike larvae which look completely different from the adults, and reach adulthood after several moultings.

Overwinter: To pass, spend or survive the winter.

Parasitize: The condition wherein one animal feeds on another living animal, often causing the death of the latter.

Photophobic: Having an aversion to light.

Phototaxis: Change in the response of an organism caused by light; generally, movement towards or away from light.

Phytophagous: Feeding on plants.

Polyphagous: Feeding on a variety of plants.

Protergum: The dorsal surface of the first body segment.

Prothorax: Anterior of the three thoracic segments.

Punctum: A depression.

Skeletonize: To reduce (a leaf) to a skeleton (by eating all the leaf tissues and leaving only leaf veins).

Sulci: Sutures formed by an enfolding of the body wall; plural of sulcus.

Suture: The junction or line of junction of adjacent parts.

Tubercle: A small, knob-like protuberance.

Verruca: A small, flat, wart-like prominence; plural – verrucae.

Vertex: The top of the head, between the eyes and anterior to the occipital suture.



APPENDIX

Check List of Bamboo Pests and Hosts

Insect Species	Main Hosts	Reported Occurrence	Pest Status
1 DEFOLIATORS			
<i>Algedonia [Pyrausta] bambucivora</i>	<i>Bambusa nutans</i> , <i>B. vulgaris</i> , <i>Dendrocalamus giganteus</i> , <i>D. strictus</i> , <i>Schizostachyum pergracile</i>	Bangladesh, India, Pakistan	Major
<i>Algedonia [Pyrausta] coclesalis</i>	<i>Arundinaria spp.</i> , <i>Bambusa vulgaris</i> , <i>Dendrocalamus giganteus</i> , <i>D. latiflorus</i> , <i>D. strictus</i> , <i>Phyllostachys pubescens</i> , <i>P. viridis</i> , <i>Schizostachyum pergracile</i>	Bangladesh, Cambodia, China, India, Indonesia, Japan, Korea, Laos, Malaysia, Myanmar, Pakistan, Sri Lanka, Vietnam	Major
<i>Allobremeria plurilineata</i>	<i>Phyllostachys pubescens</i>	China	Major
<i>Balateae gracilis</i>	<i>Phyllostachys spp.</i>	China	Minor
<i>Besaia goddrica</i>	<i>Phyllostachys spp.</i>	China	Not reported
<i>Brtona intermediana</i>	<i>Phyllostachys spp.</i>	China	Minor
<i>Brtona funeralis</i>	<i>Arundinaria spp.</i> , <i>Bambusa spp.</i> , <i>Phyllostachys spp.</i> , <i>Pseudosasa spp.</i>	China, India, Japan, Korea	Major
<i>Calaenorrhinus ambareesa</i>	<i>Bambusa bambos</i>	India	Minor
<i>Calmochrous pentasaris</i>	<i>Dendrocalamus strictus</i>	Bangladesh, India	Not reported
<i>Ceracris kiangsu</i>	<i>Phyllostachys pubescens</i>	China	Major
<i>Ceracris nigricornis</i>	<i>Phyllostachys pubescens</i>	China	Minor
<i>Ceracris nigricornis laeta</i>	<i>Phyllostachys pubescens</i>	China	Minor



Insect Species	Main Hosts	Reported Occurrence	Pest Status
<i>Circobotys aurealis</i>	<i>Bambusa</i> spp., <i>Phyllostachys</i> spp., <i>Pleioblastus</i> spp.	China, Japan, Korea, Myanmar	Not reported
<i>Cosmopterix phyllostachysae</i>	<i>Phyllostachys pubescens</i>	China, Japan	Not reported
<i>Cosmotriches</i> sp.	<i>Bambusa textilis</i> , <i>Phyllostachys</i> spp.	China	Not reported
<i>Crocidophora evenoralis</i>	<i>Bambusa</i> spp., <i>Phyllostachys</i> spp., <i>Pleioblastus</i> spp.	China, Japan, Korea, Myanmar	Not reported
<i>Crocidophora ptyophora</i>	—	India, Myanmar	Minor
<i>Demobotys pervulgaris</i>	<i>Phyllostachys pubescens</i>	China	Minor
<i>Eumorphobotys obscuralis</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Phyllostachys</i> spp.	China, Japan	Not reported
<i>Eumorphobotys eumorphalis</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Phyllostachys</i> spp.	China, Japan	Not reported
<i>Eustotia</i> sp.	—	China	Minor
<i>Eutomostethus nigrinus</i>	<i>Phyllostachys</i> spp.	China	Major
<i>Glyphipterix semiflavana</i>	<i>Phyllostachys</i> spp.	China, Japan	Not reported
<i>Hieroglyphus banian</i>	<i>Dendrocalamus strictus</i>	India	Minor
<i>Hieroglyphus tonkinensis</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Sinobambusa</i> spp., <i>Lingnania</i> spp.	China	Major
<i>Kirinia epaminondes</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Latoria parapuneta</i>	—	China	Minor
<i>Leptispa godwini</i>	<i>Dendrocalamus</i> spp., <i>Phyllostachys</i> spp., <i>Pleioblastus</i> spp., <i>Sinocalamus</i> spp.	China	Not reported



Insect Species	Main Hosts	Reported Occurrence	Pest Status
<i>Leptispa longipennis</i>	<i>Bambusa pervariabilis</i> , <i>Phyllostachys</i> spp.	China	Not reported
<i>Leptispa mevidiana</i>	<i>Bambusa pervariabilis</i> , <i>Phyllostachys</i> spp.	China	Not reported
<i>Lethe diana</i>	—	China	Not reported
<i>Lethe drypetis todara</i>	<i>Bambusa</i> spp.	India	Minor
<i>Lethe europa</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Lethe incana</i>	<i>Drepanostachyum falcatum</i>	India	Minor
<i>Lethe syrcis</i>	—	China	Not reported
<i>Lethe verma</i>	<i>Drepanostachyum falcatum</i>	India	Minor
<i>Lethe yama</i>	<i>Drepanostachyum falcatum</i>	India	Minor
<i>Liccana terminicana</i>	—	China	Not reported
<i>Lichacodia idiostygia</i>	—	China	Not reported
<i>Lichacodia squalida</i>	—	China	Not reported
<i>Loudonta dispar</i>	<i>Phyllostachys</i> spp., <i>Pleioblastus</i> spp.	China	Major
<i>Massepha absolutalis</i>	<i>Dendrocalamus strictus</i>	Bangladesh, India, Sri Lanka	Minor
<i>Matapa aria</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Phyllostachys</i> spp.	China	
<i>Microstaga jessica</i>	<i>Phyllostachys edulis</i> , <i>P. nigra</i>	Japan, Korea	Not reported
<i>Mimopydna insignis</i>	—	China	Not reported
<i>Mycalesis gotoma</i>	—	China	Not reported
<i>Neope muirheadi</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Niganda strigifascia</i>	<i>Arundinaria</i> spp., <i>Phyllostachys</i> spp.	Bhutan, China, India, Indonesia, Sikkim	Not reported
<i>Norraca decurrens</i>	—	China, India, Vietnam	Not reported



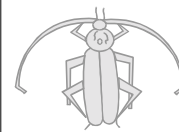
Insect Species	Main Hosts	Reported Occurrence	Pest Status
<i>Norraca retrofusca</i>	—	China, India, Vietnam	Not reported
<i>Onryza</i> sp.	<i>Phyllostachys</i> spp.	China	Not reported
<i>Oraura ordgara</i>	—	China, the Philippines	Not reported
<i>Pantana phyllostachysae</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Pantana pluto</i>	<i>Phyllostachys pubescens</i>	China, India, Indonesia, Myanmar, Vietnam	Not reported
<i>Pantana sinica</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Parasa bicolor</i>	<i>Arundinaria</i> spp., <i>Bambusa</i> spp., <i>Phyllostachys</i> spp., <i>Pleioblastus</i> spp., <i>Sinobambusa</i> spp.,	China, India, Myanmar, Nepal, Sikkim, Sri Lanka	Minor
<i>Paraplesia adelma</i>	—	China	Not reported
<i>Parnara ganga</i>	<i>Fargesia spathacea</i> , <i>Phyllostachys</i> spp.	China	Not reported
<i>Parnara guttata</i>	<i>Fargesia spathacea</i> , <i>Phyllostachys</i> spp.	China	Not reported
<i>Pelopidas agna</i>	<i>Fargesia spathacea</i> , <i>Phyllostachys</i> spp.	China	Not reported
<i>Pelopidas mathias</i>	<i>Fargesia spathacea</i> , <i>Phyllostachys</i> spp.	China	Not reported
<i>Pionea flavofimbriata</i>	<i>Dendrocalamus strictus</i>	India, Sri Lanka	Minor
<i>Philudoria albomaculata</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Philudoria divisa sulphurea</i>	—	China	Not reported
<i>Philudoria potatoris</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Phlaeoba angustidorsis</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Poecilocerus pictus</i>	<i>Dendrocalamus strictus</i>	India, Pakistan	Minor
<i>Potanthus confucius</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Rivula biatomea</i>	<i>Phyllostachys</i> spp., <i>Sinocalamus oldhami</i>	Taiwan-China	Not reported



Insect Species	Main Hosts	Reported Occurrence	Pest Status
<i>Schistocera gregaria</i>	<i>Dendrocalamus strictus</i>	India	Minor
<i>Stenadonta radialis</i>	<i>Dendrocalamus latiflorus</i> , <i>Phyllostachys</i> spp.	China, India	Minor
<i>Sylepta derogata</i>	—	Bangladesh, India, Pakistan	Not reported

2 SAP SUCKERS

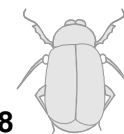
<i>Ablemothrips</i> sp.	—	India	Minor
<i>Acanthocoris scaber</i>	—	China, India, Malaysia, Myanmar, Sri Lanka	Not reported
<i>Aenaria pinchii</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Acestra sinica</i>	—	China	Not reported
<i>Agriothrips</i> sp.	—	India	Minor
<i>Aleurocanthus arundinacea</i>	<i>Bambusa bambos</i>	India	Minor
<i>Aleurocanthus bambusae</i>	<i>Bambusa nana</i> , <i>Dendrocalamus giganteus</i>	India	Minor
<i>Aleurocanthus longispinus</i>	<i>Bambusa nana</i> , <i>Dendrocalamus giganteus</i>	India	Minor
<i>Aleurocanthus niger</i>	<i>Bambusa nana</i> , <i>Dendrocalamus giganteus</i>	India	Minor
<i>Aleurocanthus nigricollis</i>	<i>Bambusa nana</i> , <i>Dendrocalamus giganteus</i>	India	Minor
<i>Aleurocanthus obovalis</i>	<i>Bambusa nana</i> , <i>Dendrocalamus giganteus</i>	India	Minor
<i>Allorothrips</i> spp.	—	India	Minor
<i>Androthrips coimbatorensis</i>	<i>Bambusa bambos</i>	India	Minor
<i>Antilliothrips</i> spp.	—	India	Minor
<i>Antonina crawii</i>	—	China	Not reported
<i>Antonina indica</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Phyllostachys</i> spp., <i>Sinocalamus</i> spp.	China, India, Japan, Sri Lanka,	Not reported



Insect Species	Main Hosts	Reported Occurrence	Pest Status
<i>Antonina pretiosa</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Phyllostachys</i> spp., <i>Sinocalamus</i> spp.	China, India, Japan, Sri Lanka,	Not reported
<i>Antonina zonata</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Phyllostachys</i> spp., <i>Sinocalamus</i> spp.	China, India, Japan, Sri Lanka,	Not reported
<i>Aphrophora horizontalis</i>	<i>Phyllostachys pubescens</i>	China	Not reported
<i>Aponychus corpuzae</i>	<i>Phyllostachys</i> spp.	China, Japan	Minor
<i>Astegopteryx bambusifoliae</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Phyllostachys</i> spp., <i>Sinocalamus</i> spp.	China, Japan, Southeast Asian countries	Not reported
<i>Astegopteryx insularis</i>	—	China, Indonesia, Japan, Malaysia	Not reported
<i>Asterolecanium bambusae</i>	<i>Bambusa vulgaris</i>	China, India, the Philippines	Major
<i>Asterolecanium coronatum</i>	<i>Bambusa</i> spp., <i>Dendrocalamus giganteus</i>	India	Minor
<i>Asterolecanium delicatum</i>	<i>Bambusa bambos</i>	India	Minor
<i>Asterolecanium flavoalatum</i>	<i>Bambusa bambos</i>	India	Minor
<i>Asterolecanium lanceolatum</i>	<i>Bambusa</i> spp., <i>Dendrocalamus giganteus</i>	India	Minor
<i>Asterolecanium logum</i>	<i>Bambusa</i> spp., <i>Dendrocalamus giganteus</i>	India	Minor
<i>Asterolecanium pudibundum</i>	—	India	Not reported
<i>Asterolecanium rubronumatum</i>	—	India	Not reported
<i>Asterolecanium soleonophoides</i>	—	India	Not reported
<i>Asterolecanium udagammae</i>	—	India	Not reported



Insect Species	Main Hosts	Reported Occurrence	Pest Status
<i>Baliothrips</i> spp.	—	India	Minor
<i>Bambusaspis bambusae</i>	<i>Bambusa</i> spp.	China, India	Minor
<i>Bambusaspis hemisphaerica</i>	<i>Arundinaria chino</i> , <i>Bambusa metake</i> , <i>Phyllostachys</i> spp.	China, Japan	Not reported
<i>Bambusaspis masuui</i>	—	China, India, Japan, the Philippines, Sri Lanka	Minor
<i>Bambusaspis miliaris</i>	—	China, India, Japan, the Philippines, Sri Lanka	Minor
<i>Bamboosiella bicoloripes</i>	<i>Bambusa bambos</i>	India	Minor
<i>Bambusiphaga</i> spp.	—	China	Minor
<i>Beloceras</i> spp.	—	China	Minor
<i>Bolacidothrips</i> sp.	—	India	Minor
<i>Brachymna tanuis</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Callitettix braconoides</i>	—	China	Minor
<i>Ceratovacuna</i> spp.	—	China, Indonesia, Japan, Malaysia	Not reported
<i>Chaetococcus bambusae</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Phyllostachys</i> spp., <i>Sinocalamus</i> spp.	China, India, Japan, Sri Lanka,	Not reported
<i>Chionaspis arundinaria</i>	<i>Phyllostachys</i> spp.	China, India	Minor
<i>Chionaspis bambusae</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Chionaspis elongata</i>	—	India	Minor
<i>Chionaspis longissima</i>	—	India	Minor
<i>Chionaspis spiculata</i>	<i>Phyllostachys</i> spp.	China, India	Minor
<i>Chucallis bambusicola</i>	<i>Miscenthus</i> sp., <i>Sinocalamus oldhami</i> , <i>Dendrocalamus latiflorus</i>	China	Minor
<i>Cloresmus modestus</i>	—	China	Not reported



Insect Species	Main Hosts	Reported Occurrence	Pest Status
<i>Cloesmus pulchellus</i>	—	China	Not reported
<i>Cosmoscarta mandarina</i>	—	China	Minor
<i>Cressona divaricata</i>	—	China	Not reported
<i>Cressona valida</i>	—	China	Not reported
<i>Ctenidiotrips</i> sp.	—	India	Minor
<i>Ctenothrips</i> sp.	—	India	Minor
<i>Derepteryx fuliginosa</i>	—	China, Japan, Korea	Not reported
<i>Dorcadotrips</i> sp.	—	India	Minor
<i>Epeuryssa</i> spp.	—	China	Minor
<i>Eriococcus rugosus</i>	<i>Phyllostachys pubescens</i>	China	Not reported
<i>Eriococcus transversus</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Phyllostachys</i> spp., <i>Sinocalamus</i> spp.	China, India, Japan, Sri Lanka,	Not reported
<i>Fracastorius cornutus</i>	—	China	Not reported
<i>Greenaspis bambusifoliae</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Greenaspis elongata</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Halyabbas unicolor</i>	--	China	Not reported
<i>Haplothrips</i> sp.	—	India	Minor
<i>Heliococcus bambusaniae</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Phyllostachys</i> spp., <i>Sinocalamus</i> spp.	China, India, Japan, Sri Lanka,	Not reported
<i>Helionothrips</i> sp.	—	India	Minor
<i>Hippotiscus dorsalis</i>	<i>Phyllostachys</i> spp.	China	Major
<i>Karnyothrips</i> spp.	—	India	Minor
<i>Kiritshenkella magnetubulata</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Phyllostachys</i> spp., <i>Sinocalamus</i> spp.	China, India, Japan, Sri Lanka,	Not reported



Insect Species	Main Hosts	Reported Occurrence	Pest Status
<i>Kuwanaspis howarai</i>	<i>Phyllostachys</i> spp.	China	Major
<i>Kuwanaspis pseudoleucaspis</i>	<i>Phyllostachys</i> spp.	China	Major
<i>Macropes</i> spp.	—	China	Not reported
<i>Malcus setosus</i>	—	China	Not reported
<i>Mecistoscolis scitetooides</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Phyllostachys</i> spp., <i>Sinocalamus</i> spp.	China, Myanmar, Sri Lanka, Taiwan-China	Major
<i>Megarrhamphus hastatus</i> —	—	China, India, Japan, Korea, Malaysia, Myanmar, Vietnam	Not reported
<i>Megarrhamphus truncatus</i>	—	China, India, Japan, Korea, Malaysia, Myanmar, Vietnam	Not reported
<i>Mystilus priamus</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Phyllostachys</i> spp., <i>Sinocalamus</i> spp.	China, Myanmar, Sri Lanka	Not reported
<i>Nesticoccus sinensis</i>	<i>Arundinaria</i> spp., <i>Phyllostachys</i> spp.	China	Not reported
<i>Notobitus excellens</i>	—	China	Not reported
<i>Notobitus meleagris</i>	<i>Dendrocalamus</i> spp.	China, India, Myanmar, Singapore, Vietnam	Not reported
<i>Notobitus montanus</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Notobitus sexguttatus</i>	<i>Dendrocalamus</i> spp.	China	Not reported
<i>Odonaspis canaliculatus</i>	<i>Bambusa vulgaris</i> , <i>Phyllostachys</i> spp.	China, India	Minor
<i>Odonaspis elongata</i>	<i>Bambusa vulgaris</i> , <i>Phyllostachys</i> spp.	China, India	Minor
<i>Odonaspis inusittatus</i>	<i>Bambusa vulgaris</i> , <i>Phyllostachys</i> spp.	China, India	Minor
<i>Odonaspis penicillata</i>	<i>Bambusa vulgaris</i> , <i>Phyllostachys</i> spp.	China, India	Minor



Insect Species	Main Hosts	Reported Occurrence	Pest Status
<i>Odonaspis senireta</i>	<i>Bambusa vulgaris</i> , <i>Phyllostachys</i> spp.	China, India	Minor
<i>Odonaspis simplex</i>	<i>Bambusa vulgaris</i> , <i>Phyllostachys</i> spp.	China, India	Minor
<i>Oligonychus sachari</i>	—	India	Minor
<i>Oligonychus uruma</i>	—	Taiwan-China	Not reported
<i>Oregma bambusae</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp.	Bangladesh, China, India, Pakistan, Sri Lanka	Major
<i>Pirkimerus japonicus</i>	<i>Phyllostachys</i> spp.	China, Japan	Not reported
<i>Podothrips bicolor</i>	<i>Bambusa bambos</i>	India	Minor
<i>Praepodothrips</i> sp.	—	India	Minor
<i>Pseudoregmasp.</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Gigantochloa</i> spp., <i>Thyrsostachys</i> spp.	Thailand	Not reported
<i>Pseudoregma albostriata</i>	<i>Miscenthus</i> sp., <i>Sinocalamus oldhami</i> , <i>Dendrocalamus latiflorus</i>	China	Minor
<i>Pseudoregma bambusicola</i>	—	China, Japan	Not reported
<i>Pseudoregma alexanderi</i>	<i>Miscenthus</i> sp., <i>Sinocalamus oldhami</i> , <i>Dendrocalamus latiflorus</i>	China	Minor
<i>Purohita</i> spp.	—	China, India	Minor
<i>Riptortus linearis</i>	—	China, India, Malaysia, Myanmar, Sri Lanka	Not reported
<i>Riptortus pedestris</i>	—	China, India, Malaysia, Myanmar, Sri Lanka	Not reported
<i>Schizotetranychus bambusae</i>	<i>Phyllostachys</i> spp.	China	Major
<i>Schizotetranychus celarus</i>	—	Japan	Minor
<i>Schizotetranychus floresi</i>	—	The Philippines	Not reported



Insect Species	Main Hosts	Reported Occurrence	Pest Status
<i>Schizotetranychus nanjingensis</i>	<i>Phyllostachys</i> spp.	China	Major
<i>Stephanothrips</i> sp.	—	India	Minor
<i>Stigmothrips</i> sp.	—	India	Minor
<i>Takecallis arundinariae</i>	—	China, Indonesia, Japan, Malaysia	Not reported
<i>Takecallis taiwanus</i>	—	China, Indonesia, Japan, Malaysia	Not reported
<i>Trionymus pulverarius</i>	<i>Bambusa</i> spp., <i>Dendrocalamus</i> spp., <i>Phyllostachys</i> spp., <i>Sinocalamus</i> spp.	China, India, Japan, Sri Lanka	Not reported
<i>Unachionaspis bambusae</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Unachionaspis signata</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Veerabahuthrips bambusae</i>	<i>Bambusa bambos</i>	India	Minor

3 SHOOT AND CULM BORERS

<i>Acanthonevra formosana</i>	—	China	Not reported
<i>Acrocratitis plumosa</i>	—	China	Not reported
<i>Apamea cuneata</i>	<i>Phyllostachys</i> spp.	China	Minor
<i>Apamea kumaso</i>	<i>Phyllostachys</i> spp.	China	Minor
<i>Apamea repetita conjuncta</i>	<i>Phyllostachys</i> spp.	China	Minor
<i>Cyrtotrachelus buqueti</i>	<i>Bambusa pervariabilis</i> , <i>B. textilis</i> , <i>Lingnania chungii</i> , <i>Sinocalamus oldhami</i>	China	Major
<i>Cyrtotrachelus dux</i>	<i>Bambusa pervariabilis</i> , <i>B. textilis</i> , <i>Dendrocalamus hamiltonii</i> , <i>D. strictus</i> , <i>Lingnania chungii</i> , <i>Melocanna baccifera</i> , <i>Sinocalamus oldhami</i>	Bangladesh, China, India, Myanmar, Sri Lanka	Major



Insect Species	Main Hosts	Reported Occurrence	Pest Status
<i>Cyrtotrachelus longimanus</i>	<i>Bambusa pervariabilis</i> , <i>B. textilis</i> , <i>Dendrocalamus hamiltonii</i> , <i>D. strictus</i> , <i>Lingnania chungii</i> , <i>Melocanna baccifera</i> , <i>Sinocalamus oldhami</i>	Bangladesh, China, India, Myanmar, Sri Lanka	Major
<i>Chelyophora ceratitina</i>	<i>Dendrocalamus strictus</i>	Bangladesh, India	Minor
<i>Chelyophora striata</i>	<i>Dendrocalamus strictus</i>	India, Sri Lanka	Minor
<i>Dichromia claripennis</i>	<i>Phyllostachys</i> spp.	China	Minor
<i>Epiparbattia gloriosalis</i>	<i>Sinocalamus affinis</i>	China	Not reported
<i>Estigmena chinensis</i>	<i>Bambusa bambos</i> , <i>B. burmicana</i> , <i>B. nutans</i> , <i>Dendrocalamus strictus</i> , <i>Gigantochloa scortechinii</i> , <i>Schizostachyum pergracile</i>	Bangladesh, India, Myanmar, Malaysia	Major
<i>Gastrozona maccuarti</i>	—	China	Not reported
<i>Myocalandra exarata</i>	<i>Bambusa polymorpha</i> , <i>Dendrocalamus strictus</i>	India	Minor
<i>Olethreutes paragramma</i>	<i>Dendrocalamus strictus</i>	Bangladesh, India, Pakistan	Major
<i>Oligia apameoidis</i>	<i>Phyllostachys</i> spp.	China	Major
<i>Oligia vulgaris</i>	<i>Phyllostachys</i> spp.	China, India, Japan	Major
<i>Oligia vulnerata</i>	<i>Phyllostachys</i> spp.	China	Major
<i>Otidognathus davidis</i>	<i>Indocalamus</i> spp., <i>Phyllostachys</i> spp., <i>Pleioblastus</i> spp., <i>Pseudosasa</i> spp., <i>Semiarundinaria</i> spp., <i>Sinobambusa</i> spp.	China, Vietnam, Southeast Asian countries	Major
<i>Otidognathus nigropictus</i>	<i>Indocalamus</i> spp., <i>Phyllostachys</i> spp., <i>Pleioblastus</i> spp., <i>Pseudosasa</i> spp., <i>Semiarundinaria</i> spp., <i>Sinobambusa</i> spp.	China, Vietnam, Southeast Asian countries	Minor



Insect Species	Main Hosts	Reported Occurrence	Pest Status
<i>Otidognathus rubriceps</i>	<i>Indocalamus</i> spp., <i>Phyllostachys</i> spp., <i>Pleioblastus</i> spp., <i>Pseudosasa</i> spp., <i>Semiarundinaria</i> spp., <i>Sinobambusa</i> spp.	China, Vietnam, Southeast Asian countries	Minor
<i>Parahyrataea jacobsonae</i>	—	China	Not reported
<i>Pegomyia kiangsuensis</i>	<i>Phyllostachys</i> spp.	China	Minor
<i>Pegomyia phyllostachys</i>	<i>Phyllostachys</i> spp.	China	Minor
<i>Pterolophia trilineicollis</i>	—	China	Not reported
<i>Ptilona persimilis</i>	—	China	Not reported
<i>Purpuricenus temminckii</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Sagra femorata purpurea</i>	—	China, Vietnam	Minor

4 GALL MAKERS

<i>Aiolomorphus rhopaloides</i>	<i>Phyllostachys</i> spp.	China, Japan	Major
<i>Ceraphron</i> sp.	<i>Bambusa bambos</i>	India	Not reported
<i>Diomorus aioromorthi</i>	<i>Phyllostachys</i> spp.	China, Japan	Not reported
<i>Planetella conesta</i>	—	China	Not reported

5 RHIZOME AND ROOT FEEDERS

<i>Agonischius obscuripes</i>	—	China	Not reported
<i>Agriotes</i> sp.	—	China	Not reported
<i>Ceylonitermes indicola</i>	—	India	Minor
<i>Chyliza bambusae</i>	<i>Phyllostachys pubescens</i>	China	Not reported
<i>Coptotermes heimi</i>	—	India	Minor
<i>Dorystenes buqueti</i>	—	China, India, Indonesia, Laos, Malaysia, Myanmar, Nepal	Minor



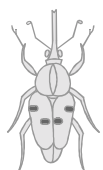
Insect Species	Main Hosts	Reported Occurrence	Pest Status
<i>Dorysthenes walkeri</i>	—	China, India, Myanmar	Minor
<i>Holotrichia</i> sp.	<i>Bambusa bambos</i> , <i>B. nutans</i> , <i>Dendrocalamus strictus</i>	India	Minor
<i>Huechys sanguinea</i>	—	China	Not reported
<i>Lacon</i> sp.	—	China	Not reported
<i>Melanotus regalis</i>	—	China	Not reported
<i>Microcerotermes fletcheri</i>	—	India	Minor
<i>Microcerotermes heimi</i>	—	India	Minor
<i>Microcerotermes obesi</i>	—	India	Minor
<i>Mogannia hebes</i>	—	China	Not reported
<i>Platylomia pieli</i>	<i>Phyllostachys</i> spp.	China	Not reported
<i>Pseudocapritermes goanicus</i>	—	India	Minor

6 SEED PESTS

<i>Sitotroga cereallela</i>	<i>Bambusa burmicana</i> , <i>B. tulda</i>	Bangladesh, India	Minor
<i>Udonga montana</i>	<i>Bambusa polymorpha</i> , <i>Dendrocalamus strictus</i> , <i>Melocanna baccifera</i>	Bangladesh, India, Myanmar	Major

7 NURSERY PESTS

<i>Hieroglyphus banian</i>	—	India, Pakistan	Minor
<i>Holotrichia</i> sp.	<i>Dendrocalamus strictus</i>	India	Minor
<i>Odontotermes</i> sp.	<i>Bambusa tulda</i>	India	Minor
<i>Poecilocerus pictus</i>	—	India, Pakistan	Minor



Insect Species	Main Hosts	Reported Occurrence	Pest Status
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8 POST-HARVEST PESTS

<i>Ceresium sinicum</i>	—	China, Japan, Thailand	Major
<i>Chlorophorus annularis</i>	<i>Bambusa</i> spp., <i>Dendrocalamus strictus</i> <i>Phyllostachys</i> spp.	China, India, Indonesia, Japan, Malaysia, Myanmar, Thailand, Vietnam	Major
<i>Chlorophorus figuratue</i>	—	China	Not reported
<i>Coptotermes heimi</i>	—	India, Pakistan	Not reported
<i>Cryphalus satonis</i>	—	China	Not reported
<i>Cryptotermes dudleyi</i>	—	Bangladesh, India	Major
<i>Diboma posticata</i>	—	China	Not reported
<i>Dinoderus brevis</i>	<i>Bambusa bambos</i> , <i>B. pervariabilis</i> , <i>B. polymorpha</i> , <i>B. textilis</i> , <i>B. vulgaris</i> , <i>Dendrocalamus giganteus</i> , <i>D. hamiltonii</i> , <i>D. strictus</i> , <i>Phyllostachys pubescens</i>	Bangladesh, China, India,	Major Pakistan
<i>Dinoderus japonicus</i>	<i>Phyllostachys viridis</i> , <i>P. pubescens</i> , <i>Pleiolblastus amarus</i>	China, Japan, most bamboo-growing countries	Major
<i>Dinoderus minutus</i>	<i>Bambusa bambos</i> , <i>B. pervariabilis</i> , <i>B. polymorpha</i> , <i>B. textilis</i> , <i>B. vulgaris</i> , <i>Dendrocalamus giganteus</i> , <i>D. hamiltonii</i> , <i>D. strictus</i> , <i>Phyllostachys pubescens</i>	Bangladesh, China, India,	Major Pakistan
<i>Dinoderus ocellaris</i>	<i>Bambusa bambos</i> , <i>B. pervariabilis</i> , <i>B. polymorpha</i> , <i>B. textilis</i> , <i>B. vulgaris</i> , <i>Dendrocalamus giganteus</i> , <i>D. hamiltonii</i> , <i>D. strictus</i> , <i>Phyllostachys pubescens</i>	Bangladesh, China, India,	Major Pakistan
<i>Dinoderus pilifrons</i>	—	China	Not reported
<i>Glenea notata</i>	—	China	Not reported



Insect Species	Main Hosts	Reported Occurrence	Pest Status
<i>Heterobostrychus aequalis</i>	—	—	India Minor
<i>Heterobostrychus hamatipennis</i>	—	China	Not reported
<i>Lyctus africanus</i>	<i>Bambusa bambos</i> , <i>Dendrocalamus strictus</i>	India	Minor
<i>Lyctus brunneus</i>	—	Southeast Asian countries	Minor
<i>Lyctus sinensis</i>	—	China	Not reported
<i>Merionoeda nigriceps</i>	—	China	Not reported
<i>Microcerotermes fletcheri</i>	—	India	Not reported
<i>Microcerotermes heimi</i>	—	India	Not reported
<i>Minthea rugicollis</i>	—	Tropical and subtropical countries	Not reported
<i>Niphona furcata</i>	<i>Phyllostachys</i> spp., <i>Pleioblastus</i> spp.	China, Japan	Not reported
<i>Niphona hookeri</i>	—	India	Not reported
<i>Purpuricenus sanguinolentus</i>	—	China	Not reported
<i>Ropica signata</i>	—	China	Not reported
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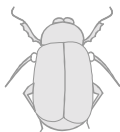
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